

## Traditional Medicine Combination Therapy Is A Promising Strategy for MRSA Infection

Table 1. MRSA virulence factors that contribute to immune evasion and their functions in host immune responses

Evasion proteins against mucosal immune	Target	Function/ effect on immune system	Ref.
<i>Staphylococcal</i> superantigen-like protein-7 (SSL-7)	SIgA	It enhances the ability to colonize in mucosal environments such as the nasal passage by binding to SIgA.	[1]
Evasion proteins against innate immune	Target	Function/ effect on immune system	Ref.
Proteins including extracellular adherence protein (Eap), collagen-binding protein (Cna) and serine-aspartate repeat protein E (SdrE)	C1q in classical pathway, lectin pathway, alternative pathway	They inhibit the activation of the complement system by blocking these three pathways.	[2-4]
<i>Staphylococcal</i> Protein A (SpA)	IgG	It results in the inverted tagging and blocking of the C1q binding sites, preventing complement initiation.	[5]
	B cells	It interferes in B-cells activation and proliferation, reducing the phagocytosis of MRSA, impeding antibody production and causing disordered activation, finally leading to the death of the B cells.	[6, 7]

<i>Staphylococcal</i> complement inhibitor (SCIN)	C3 convertases	It binds and stabilizes C3 convertases, interfering with the activation of the complement system.	[8]
Extracellular fibrinogen-binding protein (Efb); its homolog extracellular complement-binding protein (Ecb)	C3	They prevent C3 from recognition by macrophages.	[9]
Second Immunoglobulin-binding protein (Sbi)	IgG	It avoids neutrophil-mediated opsonophagocytosis.	[10]
	C3	It binds to complement, leading to the cleavage and consumption of complement Factor C3.	[11]
Factor I	C3b	It mediates C3b cleavage to iC3b, inhibiting initiation of the alternative pathway as well the activation of the terminal complement cascade.	[12]
Staphylokinase (SAK)	Complement	It digests IgG and complement.	[13]
chemotaxis inhibitory protein of staphylococci (CHIPS)	C5aR	It binds to C5aR to evade complement.	[14]
	FPR1	It binds to FPR1 to block the chemotaxis of neutrophils.	[15]
<i>Staphylococcal</i> superantigen-like protein-7 (SSL-7)	C5	It inhibits the opsonization of bacteria by inhibiting the cleavage of C5.	[16]
Proteases, e.g., staphopain A (Scp A), aureolysin	Complement	They degrade complement preventing opsonization and bacteria	[5]

(Aur)		lysis.	
	Neutrophils	ScpA inhibits the chemotaxis and activation of neutrophils.	[17]
<i>Staphylococcal</i> superantigen-like 5 (SSL5)	P-selectin glycoprotein ligand 1 (PSGL-1)	It binds to PSGL-1 of sialyl Lewis X, thus blocking PSGL-1 interaction with the natural ligand P-selectin and abrogating neutrophil rolling on endothelial cells.	[18]
Proteins including extracellular adherence protein (Eap)	Intercellular adhesion molecule 1 (ICAM-1)	It binds to ICAM-1, blocking the neutrophil recruitment to the infection site.	[8]
<i>Staphylococcal</i> complement inhibitor (SCIN), chemotaxis inhibitory protein of staphylococci (CHIPS)	Neutrophils	They impair neutrophil recruitment as chemotaxis inhibitors.	[19]
Nuc	NETs	It is a nuclease produced by MRSA to evade NETs.	[20, 21]
Panton-Valentine leukocidin (PVL)	Neutrophils	It causes neutrophil lysis at the dose of 0.04 µg/mL.	[22]
Phenol-soluble-modulins (PSMs)	Neutrophils	They directly kill neutrophils mainly by disrupting the membrane.	[23]



Table 2. The effect of TM therapies on host immunity and MRSA immune evasion.

Name of TM	Active Ingredient	Model	Dosing concentration	Targets and indicators	Immunomodulatory action	Ref.
<i>G. glavra</i>	Glycyrrhizin (GL)	Male CD mice (20–25 g)	10 mg/kg	ICAM-1, P-selectin	It decreases the degree of positive staining and densitometry for ICAM-1 and P-selectin.	[24]
		Male specific pathogen-free BALB/c mice (6 weeks)	50 mg/kg	neutrophils, macrophages, MPO, CD11b	It reduces the numbers of total cells, neutrophils, and macrophages, and decreases MPO and CD11b levels to inhibit the migration and infiltration of immune cells.	[24, 25]
				TNF- $\alpha$ , IL-1 $\alpha$ , IL-6, COX-2, iNOS, NF- $\kappa$ B	It decreases the levels of pro-inflammatory cytokines (TNF- $\alpha$ , IL-1 $\alpha$ , IL-6) and pro-inflammatory mediators (COX-2, iNOS, NF- $\kappa$ B).	[24, 25]
	18 $\beta$ -Glycyrrhetic Acid (GA)	Female Crl; SKH1-hrBR hairless mice, subcutaneously inoculated with MRSA	600 $\mu$ g/mL	saeR, Hla, RNAIII, mecA, sbi	It reduces MRSA immune evasion by down-regulating MRSA virulence factors (saeR, Hla, RNAIII, mecA, sbi).	[26]
				KC, G-CSF	It reduces the levels of inflammatory cytokines (KC, G-CSF).	[26]

			co-stimulatory molecules CD40, CD86, MHC class II	It induces phenotypic maturation of DCs by increasing CD40, CD86 and MHC-II expression in DCs. [27]
	Male BALB/c and C57BL/6 mice (6 weeks)	1 and 10 mg/mL for 14 hours	IL-12, IFN- $\gamma$	It increases IL-12 levels to promote DC maturation and activates T cells to differentiate into IFN- $\gamma$ producing Th1 cells. [27]
			IL-10	It suppresses excessive inflammatory responses or terminates the immune responses after pathogen eradication by increasing IL-10 levels. [27]
<i>P. ginseng</i> (Pg)	Ginsan	Male pyrogen-free BALB/c mice (5–7 weeks, 18–22 g); <i>S. aureus</i> 25923 or <i>E. coli</i>	0.012, 0.025, 0.5, 25, and 250 mg/kg (intravenous injection)	Macrophage, TLR2, TLR4, TLR9, MyD88, MAPK, JUN 1/2, NF- $\kappa$ B, TNF- $\alpha$ , IL-1 $\beta$ , IL-6, IL-
				It induces resistance to MRSA septicemia by modulating monocyte/macrophage-mediated innate immunity. [28]

				12	
	BALB/c mice	100 mg/kg (intraperitoneally introduced, once a day)	Serum antibodies, Peyer's patches, COX-1, COX-2, CCL3	It effectively enhances the humoral immune response to orally delivered antigen, mediated by CCL3 via COX-1 and COX-2.	[29]
Pg extract, such as ginsenoside	Female mouse mastitis model (lactating mice)	3, 10, 50 mg/mL	Macrophage, TLR2, TLR4, NF-κB	Pg extracts trigger and induce the inflammatory response. Pg modulates the mRNA levels of TLR2 and TLR4, triggers the activation of the MyD88-dependent pathway and then leads to the liberation of the NF-κB transcription factor.	[30]
Ginsenoside	MRSA strains: bacterial cells cultured in a Mueller–Hinton broth	100 mg/mL	MRSA biofilm	It not only attenuates bacterial toxicity but also promotes the influx of antibiotics.	[31]
ginseng oligopeptides	420 Female healthy	0.0375, 0.075, 0.15, 0.3 and 0.6	Macrophage, NK cell, T	It increases macrophage phagocytosis capacity and NK cell activity, and	[32]

	(GOP)	BALB/c mice	g/kg for 30 days (intragastrically administered)	and Th cells	enhances T and Th cells, as well as IL-2, IL-6 and IL-12 secretion and IgA, IgG1 and IgG2b production.
<i>P. quinquefolius</i>	aqueous extract of the <i>P.</i> <i>quinquefolius</i> (CVT-E002)	Old mice (8–9 weeks)	80 mg chow/mouse daily	NK cell in spleen and bone marrow	It augments the absolute numbers of NK [33] cells in the spleen and bone marrow.
		C57 BL/6 mice	10, 100, 500 Ag/mL	B- lymphocyte in the spleen	It increases IL2 and IFN- $\gamma$ levels in B- [34] lymphocytes in a dose-dependent manner.
		C57 BL/6 mice (6–8 weeks)	500, 100 and 10 $\mu$ g/mL	Peritoneal exudate macrophages (PEM)	It stimulates NO, IL-1, IL-6 and TNF- $\alpha$ [35] levels in PEM.
		BALB/c mice (1 week)	18, 6 mg per mouse	Plasma cells	It increases IgG levels. [35]
<i>O. japonicas</i>	OPS, OPL	Peritoneal macrophages isolated from ICR mice (18– 22 g)	62.5, 31.25, 15.625, 7.813 and 3.907 g/mL	Phagocytes, NO, iNOS	It improves immune function of [36] macrophages by enhancing phagocytic function and increasing NO and iNOS levels, finally enhancing the ability of sterilization.
				IL-1 $\beta$ , TNF-	It exerts immune activity by promoting IL- [36]



					$\alpha$ , MCP-1, 1 $\beta$ , TNF- $\alpha$ , MCP-1 and MIP-1 $\beta$ levels in macrophages.	
				31.25, 15.625 and 7.813 g/mL	CD14, MHC-II	It induces CD14 and MHC-II to promote macrophages activation and maturation. [36]
				0.3, 1.0 and 3.0 mg/kg	MPO	It reduces neutrophil infiltration by decreasing MPO levels. [37]
		Male ICR mice (6–8 weeks)		1.0 and 3.0 mg/kg	iNOS, NF- $\kappa$ B	It suppresses the inflammatory response by decreasing iNOS levels, which might be linked with the down-regulation of NF- $\kappa$ B. [37]
	Ruscogenin (RUS)			0.3 and 1 mg/kg	NO, IL-6, TNF- $\alpha$	It alleviates lung injury and inflammation by decreasing NO, IL-6 and TNF- $\alpha$ levels. [38]
		Male C57BL/6 mice (18–22 g)		0.1, 0.3 and 1 mg/kg	Bax, cleaved caspase-3, Bcl-2	It inhibits PEC apoptosis by decreasing Bax and cleaved caspase-3 levels and by increasing Bcl-2 levels. [38]
				0.3 and 1 mg/kg	TLR4, MYD88, NF- $\kappa$ B p65	It attenuates LPS-induced PEC apoptosis and exerts a protective effect on lung injury and inflammation by suppressing the TLR4/MYD88/NF- $\kappa$ B pathway. [38]
Cordyceps mushrooms-	<i>Cordyceps C. sinensis</i> extract	Sixty adult	male BALB/c	10, 30 and 60 mg/kg	Neutrophils, macrophages,	It alleviates inflammatory cell exudation by decreasing the numbers of neutrophils [39]

sinensis	(CSE)	mice (8 weeks, 20 ± 2 g)	MPO NF-κB p65, COX-2, iNOS, NO, TNF-α, IL-6, IL-1β	and macrophages, as well as MPO levels. It down-regulates NO, TNF-α, IL-6 and IL-1β by inhibiting the phosphorylation of NF-κB p65 and COX-2, iNOS.	[39]
Cordyceps mushrooms- militaris	C. cordycepin	Male specific pathogen-free 1, 10 and 30 Wistar rats (8– mg/kg 10 weeks)	TNF-α, IL-6, HMGB1, IL- 10, TLR4  Neutrophils, MPO, NO, iNOS, LDH  Nrf2, HO-1	It alleviates anti-oxidative stress injuries by down regulating TNF-α, IL-6 and HMGB1 as well up-regulating IL-10, which is associated with inhibiting TLR4 signaling.  It attenuates inflammation by decreasing the neutrophil number and inhibiting their exudation by suppressing MPO, NO, iNOS and LDH levels.  It stimulates HO-1 production and alleviates lung injuries by promoting Nrf2 activation and inducing nuclear translocation of Nrf2.	[40, 41]
		Male BALB/c 50, 100 and 200 mice (6-8 mg/kg)	NF-κB, IκB	It prevents IκB phosphorylation and NF- κB release to achieve anti-inflammatory	[41]

					weeks)		effect.	
							Neutrophils, macrophages, MPO	It decreases neutrophil and macrophage numbers in BALF and inhibits neutrophil infiltration by reducing MPO levels. [42]
							TNF- $\alpha$ , IL-6, IL-1 $\beta$ , IL-13, MIF, IL-10	It decreases TNF- $\alpha$ , IL-6, IL-1 $\beta$ , IL-13 and MIF levels, as well as increases IL-10 levels in a dose-dependent manner. [42]
Atractylodes species- <i>macrocephala</i>	A.						TLR4, NF- $\kappa$ B, I $\kappa$ B $\alpha$	It exerts a protective effect on ALI-induced mice by inhibiting TLR4, NF- $\kappa$ B activation and I $\kappa$ B $\alpha$ degradation. [42]
							Macrophages, NO, TNF- $\alpha$	It stimulates macrophages to promote phagocytic activity and the productions of NO and TNF- $\alpha$ . [43, 44]
							NF- $\kappa$ B, I $\kappa$ B	It induces I $\kappa$ B degradation and the activation of NF- $\kappa$ B. [43, 44]
							Macrophages, NO, TNF- $\alpha$ , IL-6	They promote macrophages phagocytosis and the release of NO, TNF- $\alpha$ and IL-6. [45]
Atractylodes species- <i>lancea</i>	A.						HGF	They modulate the intestinal immune system by stimulating Peyer's patch cells to induce HGF production. [45]

		BALB/C mice (6-8 weeks)				
	water extracts of <i>A. macrocephala</i> and <i>A. lancea</i>	Murine normal colonic epithelial cell- line MCE301 cells	100 µg/mL	G-CSF	They promote the intestinal immune system by promoting G-CSF.	[46]
Zhenqi Fuzheng granule (ZQ)	Astragalus polysaccharides (APS)	Male MF1albino mice (5-6 weeks)	250 mg APS/kg/week for four consecutive weeks 1/mL	Spleen, neutrophil and ROS in intestinal macrophage	It increases both in the phagocytic ability of neutrophils and the intestinal ROS production. It increases the phagocytic activity of macrophages.	[47] [48]
Yu Ping Feng San (YPFS)	water-soluble extracts of YPFS	RAW 264.7 murine macrophages	1mg/mL; 3h 0.03, 0.1, 0.3, 1, 3 mg/mL; 24h 1mg/mL; 3h & 24h	IL-1β, IL-6, TNF-α IL-1β, IL-6, TNF-α iNOS, COX- 2	It suppresses the production of pro- inflammatory cytokines in LPS-induced cultured macrophages. It induces the production of pro- inflammatory cytokines. It reduces iNOS and COX-2 levels in macrophages at 3-hour time points. However, it induces the two at 24-hour	[48] [48] [49]

						time points.
	Filtered solutions	SMS	Healthy specific pathogen-free ICR mice (20–22 g, 6–8 weeks)	0.375, 0.75 and 1.5 mL/kg	IFN- $\gamma$ , TNF- $\alpha$ , IL-2, NF- $\kappa$ B	It inhibits excessive inflammation by regulating NF- $\kappa$ B and decreasing IFN- $\gamma$ , TNF- $\alpha$ and IL-2 levels. <sup>[50]</sup>
SMS					MPO, neutrophils, macrophages	It improves inflammatory cell infiltration in pulmonary tissue by inhibiting neutrophils and macrophages, as well as MPO levels. <sup>[51]</sup>
	Schisantherin A		Male BALB/c mice, (6–8 weeks)	10, 20 and 40 mg/kg	TNF- $\alpha$ , IL-6, IL-1 $\beta$ in the BALF	It exerts an anti-inflammatory effect through decreasing TNF- $\alpha$ , IL-6 and IL-1 $\beta$ levels in the BALF. <sup>[51]</sup>
					NF- $\kappa$ B p65, I $\kappa$ B- $\alpha$ , JNK, ERK, p38	It inhibits the phosphorylation of p65, I $\kappa$ B- $\alpha$ in a dose-dependent manner. <sup>[51]</sup>
Buzhongyiqitang (Hochuekkito)	Hochuekkito extract (HET)		Female BALB/c mice (6 weeks)	3.4 g/kg/day	MRSA, splenocyte	It inhibits MRSA and promotes murine splenocyte immunological activity in dose-dependent manners. <sup>[52]</sup>
			73 acute stroke	7.5 g/day, three	serum	It improves levels of serum nutritional <sup>[53]</sup>

patients (41 HET-treated and 32 non-HET-treated)	divided doses for three months	nutritional markers	markers by supporting nutrition and enhancing innate immunity.	
Female C3H/HeJ mice (6-8 weeks)	1000 mg/kg/day	mucosal IgA antibody	It enhances the IgA immune response.	[54]

Table 3. Differences in the mechanisms of the two therapies

Conventional Therapies			TM Immune Therapies
Mucosal immune response		Oral vaccines have not been developed.	They enhance Peyer's patches activation and increase the production of IgA+B cells and SIgA.
Innate immune response		<p>(a) The development and research of vaccines are mainly based on inhibiting the bacterial immune evasion from innate immune responses, for example, targeting virulence factors by blocking the complement system and killing neutrophils.</p> <p>(b) AAC therapy facilitates bacterial uptake by phagocytic cells through opsonizing by antibodies.</p>	<p>(a) Targeting the complement system: They influence the complement system to regulate the development of MRSA infection.</p> <p>(b) Targeting neutrophil phagocytosis: They act on neutrophil migration and recruitment, chemokine expression, macrophage activity and antigen presentation.</p>
Adaptive immune response		The development and research of passive immunotherapy also targets the molecules or components mediated immune evasion.	<p>(a) Targeting T-cell activation: They stimulate T-cell activation and enhancing T-lymphocyte proliferation.</p> <p>(b) Targeting antibodies: They increase B-cell activation and antibody titers.</p> <p>(c) Targeting MRSA immune evasion strategies: They inhibit the formation of bacterial biofilms.</p>

## References:

1. Bestebroer J, Aerts PC, Rooijackers SH, Pandey MK, Kohl J, van Strijp JA, et al. **Functional basis for complement evasion by staphylococcal superantigen-like 7.** *Cell Microbiol* 2010; 12(10):1506-1516.
2. Pietrocola G, Nobile G, Rindi S, Speziale P. **Staphylococcus aureus Manipulates Innate Immunity through Own and Host-Expressed Proteases.** *Front Cell Infect Microbiol* 2017; 7:166.
3. Sage MAG, Cranmer KD, Semeraro ML, Ma S, Galkina EV, Tran Y, et al. **A Factor H-Fc fusion protein increases complement-mediated opsonophagocytosis and killing of community associated methicillin-resistant Staphylococcus aureus.** *PLoS One* 2022; 17(3):e0265774.
4. Woehl JL, Ramyar KX, Katz BB, Walker JK, Geisbrecht BV. **The structural basis for inhibition of the classical and lectin complement pathways by S. aureus extracellular adherence protein.** *Protein Sci* 2017; 26(8):1595-1608.
5. Wojcik-Bojek U, Rozalska B, Sadowska B. **Staphylococcus aureus-A Known Opponent against Host Defense Mechanisms and Vaccine Development-Do We Still Have a Chance to Win?** *Int J Mol Sci* 2022; 23(2).
6. Falugi F, Kim HK, Missiakas DM, Schneewind O. **Role of protein A in the evasion of host adaptive immune responses by Staphylococcus aureus.** *mBio* 2013; 4(5):e00575-00513.
7. Pauli NT, Kim HK, Falugi F, Huang M, Dulac J, Henry Dunand C, et al. **Staphylococcus aureus infection induces protein A-mediated immune evasion in humans.** *J Exp Med* 2014; 211(12):2331-2339.
8. do Vale A, Cabanes D, Sousa S. **Bacterial Toxins as Pathogen Weapons Against Phagocytes.** *Front Microbiol* 2016; 7:42.
9. Kumar D, Romero Y, Schuck KN, Smalley H, Subedi B, Fleming SD. **Drivers and regulators of humoral innate immune responses to infection and cancer.** *Mol Immunol* 2020; 121:99-110.
10. Smith EJ, Visai L, Kerrigan SW, Speziale P, Foster TJ. **The Sbi protein is a multifunctional immune evasion factor of Staphylococcus aureus.** *Infect Immun* 2011; 79(9):3801-3809.
11. Mues N, Chu HW. **Out-Smarting the Host: Bacteria Maneuvering the Immune Response to Favor Their Survival.** *Front Immunol* 2020; 11:819.
12. Irmischer S, Doring N, Halder LD, Jo EAH, Kopka I, Dunker C, et al. **Kallikrein Cleaves C3 and Activates Complement.** *J Innate Immun* 2018; 10(2):94-105.
13. Teng TS, Ji AL, Ji XY, Li YZ. **Neutrophils and Immunity: From Bactericidal Action to Being Conquered.** *J Immunol Res* 2017; 2017:9671604.
14. Grousd JA, Rich HE, Alcorn JF. **Host-Pathogen Interactions in Gram-Positive Bacterial Pneumonia.** *Clin Microbiol Rev* 2019; 32(3).
15. Nasser A, Moradi M, Jazireian P, Safari H, Alizadeh-Sani M, Pourmand MR, et al. **Staphylococcus aureus versus neutrophil: Scrutiny of ancient combat.** *Microb Pathog* 2019; 131:259-269.
16. Zhao Y, van Kessel KPM, de Haas CJC, Rogers MRC, van Strijp JAG, Haas PA. **Staphylococcal superantigen-like protein 13 activates neutrophils via formyl peptide receptor 2.** *Cell Microbiol* 2018; 20(11):e12941.
17. Laarman AJ, Mijnheer G, Mootz JM, van Rooijen WJ, Ruyken M, Malone CL, et al. **Staphylococcus aureus Staphopain A inhibits CXCR2-dependent neutrophil activation and chemotaxis.** *EMBO J* 2012; 31(17):3607-3619.
18. Guerra FE, Borgogna TR, Patel DM, Sward EW, Voyich JM. **Epic Immune Battles of History: Neutrophils vs. Staphylococcus aureus.** *Front Cell Infect Microbiol* 2017; 7:286.
19. Rigby KM, DeLeo FR. **Neutrophils in innate host defense against Staphylococcus**



**aureus infections.** *Semin Immunopathol* 2012; 34(2):237-259.

20. Herzog S, Dach F, de Buhr N, Niemann S, Schlagowski J, Chaves-Moreno D, et al. **High Nuclease Activity of Long Persisting Staphylococcus aureus Isolates Within the Airways of Cystic Fibrosis Patients Protects Against NET-Mediated Killing.** *Front Immunol* 2019; 10:2552.

21. Schilcher K, Andreoni F, Uchiyama S, Ogawa T, Schuepbach RA, Zinkernagel AS. **Increased neutrophil extracellular trap-mediated Staphylococcus aureus clearance through inhibition of nuclease activity by clindamycin and immunoglobulin.** *J Infect Dis* 2014; 210(3):473-482.

22. Loffler B, Hussain M, Grundmeier M, Bruck M, Holzinger D, Varga G, et al. **Staphylococcus aureus panton-valentine leukocidin is a very potent cytotoxic factor for human neutrophils.** *PLoS Pathog* 2010; 6(1):e1000715.

23. Yao Z, Cary BP, Bingman CA, Wang C, Kreitler DF, Satyshur KA, et al. **Use of a Stereochemical Strategy To Probe the Mechanism of Phenol-Soluble Modulin alpha3 Toxicity.** *J Am Chem Soc* 2019; 141(19):7660-7664.

24. Menegazzi M, Di Paola R, Mazzon E, Genovese T, Crisafulli C, Dal Bosco M, et al. **Glycyrrhizin attenuates the development of carrageenan-induced lung injury in mice.** *Pharmacol Res* 2008; 58(1):22-31.

25. Lee SA, Lee SH, Kim JY, Lee WS. **Effects of glycyrrhizin on lipopolysaccharide-induced acute lung injury in a mouse model.** *J Thorac Dis* 2019; 11(4):1287-1302.

26. Long DR, Mead J, Hendricks JM, Hardy ME, Voyich JM. **18beta-Glycyrrhetic acid inhibits methicillin-resistant Staphylococcus aureus survival and attenuates virulence gene expression.** *Antimicrob Agents Chemother* 2013; 57(1):241-247.

27. Bordbar N, Karimi MH, Amirghofran Z. **Phenotypic and functional maturation of murine dendritic cells induced by 18 alpha- and beta-glycyrrhetic acid.** *Immunopharmacol Immunotoxicol* 2014; 36(1):52-60.

28. Ahn JY, Choi IS, Shim JY, Yun EK, Yun YS, Jeong G, et al. **The immunomodulator ginsan induces resistance to experimental sepsis by inhibiting Toll-like receptor-mediated inflammatory signals.** *Eur J Immunol* 2006; 36(1):37-45.

29. Na HS, Lim YJ, Yun YS, Kweon MN, Lee HC. **Ginsan enhances humoral antibody response to orally delivered antigen.** *Immune Netw* 2010; 10(1):5-14.

30. Silvestrini P, Beccaria C, Pereyra EAL, Renna MS, Ortega HH, Calvino LF, et al. **Intramammary inoculation of Panax ginseng plays an immunoprotective role in Staphylococcus aureus infection in a murine model.** *Res Vet Sci* 2017; 115:211-220.

31. Sung WS, Lee DG. **The combination effect of Korean red ginseng saponins with kanamycin and cefotaxime against methicillin-resistant Staphylococcus aureus.** *Biol Pharm Bull* 2008; 31(8):1614-1617.

32. He LX, Ren JW, Liu R, Chen QH, Zhao J, Wu X, et al. **Ginseng (Panax ginseng Meyer) oligopeptides regulate innate and adaptive immune responses in mice via increased macrophage phagocytosis capacity, NK cell activity and Th cells secretion.** *Food Funct* 2017; 8(10):3523-3532.

33. Miller SC, Ti L, Shan J. **Dietary supplementation with an extract of North American ginseng in adult and juvenile mice increases natural killer cells.** *Immunol Invest* 2012; 41(2):157-170.

34. Wang M, Guilbert LJ, Li J, Wu Y, Pang P, Basu TK, et al. **A proprietary extract from North American ginseng (Panax quinquefolium) enhances IL-2 and IFN-gamma productions in murine spleen cells induced by Con-A.** *Int Immunopharmacol* 2004; 4(2):311-315.

35. Wang M, Guilbert LJ, Ling L, Li J, Wu Y, Xu S, et al. **Immunomodulating activity of CVT-E002, a proprietary extract from North American ginseng (Panax quinquefolium).**

*J Pharm Pharmacol* 2001; 53(11):1515-1523.

36. Sun W, Hu W, Meng K, Yang L, Zhang W, Song X, et al. **Activation of macrophages by the ophiopogon polysaccharide liposome from the root tuber of *Ophiopogon japonicus*.** *Int J Biol Macromol* 2016; 91:918-925.

37. Sun Q, Chen L, Gao M, Jiang W, Shao F, Li J, et al. **Ruscogenin inhibits lipopolysaccharide-induced acute lung injury in mice: involvement of tissue factor, inducible NO synthase and nuclear factor (NF)-kappaB.** *Int Immunopharmacol* 2012; 12(1):88-93.

38. Wu Y, Wang Y, Gong S, Tang J, Zhang J, Li F, et al. **Ruscogenin alleviates LPS-induced pulmonary endothelial cell apoptosis by suppressing TLR4 signaling.** *Biomed Pharmacother* 2020; 125:109868.

39. Fu S, Lu W, Yu W, Hu J. **Protective effect of *Cordyceps sinensis* extract on lipopolysaccharide-induced acute lung injury in mice.** *Biosci Rep* 2019; 39(6).

40. Qing R, Huang Z, Tang Y, Xiang Q, Yang F. **Cordycepin alleviates lipopolysaccharide-induced acute lung injury via Nrf2/HO-1 pathway.** *Int Immunopharmacol* 2018; 60:18-25.

41. Lei J, Wei Y, Song P, Li Y, Zhang T, Feng Q, et al. **Cordycepin inhibits LPS-induced acute lung injury by inhibiting inflammation and oxidative stress.** *Eur J Pharmacol* 2018; 818:110-114.

42. Zhang JL, Huang WM, Zeng QY. **Atractylenolide I protects mice from lipopolysaccharide-induced acute lung injury.** *Eur J Pharmacol* 2015; 765:94-99.

43. Ji GQ, Chen RQ, Zheng JX. **Macrophage activation by polysaccharides from *Atractylodes macrocephala* Koidz through the nuclear factor-kappaB pathway.** *Pharm Biol* 2015; 53(4):512-517.

44. Li BX, Li WY, Tian YB, Guo SX, Huang YM, Xu DN, et al. **Polysaccharide of *Atractylodes macrocephala* Koidz Enhances Cytokine Secretion by Stimulating the TLR4-MyD88-NF-kappaB Signaling Pathway in the Mouse Spleen.** *J Med Food* 2019; 22(9):937-943.

45. Qin J, Wang HY, Zhuang D, Meng FC, Zhang X, Huang H, et al. **Structural characterization and immunoregulatory activity of two polysaccharides from the rhizomes of *Atractylodes lancea* (Thunb.) DC.** *Int J Biol Macromol* 2019; 136:341-351.

46. Shimato Y, Ota M, Asai K, Atsumi T, Tabuchi Y, Makino T. **Comparison of byakujutsu (*Atractylodes rhizome*) and sojutsu (*Atractylodes lancea* rhizome) on anti-inflammatory and immunostimulative effects in vitro.** *J Nat Med* 2018; 72(1):192-201.

47. Abuelsaad AS. **Supplementation with *Astragalus* polysaccharides alters *Aeromonas*-induced tissue-specific cellular immune response.** *Microb Pathog* 2014; 66:48-56.

48. Du CY, Choi RC, Zheng KY, Dong TT, Lau DT, Tsim KW. **Yu Ping Feng San, an ancient Chinese herbal decoction containing *Astragali Radix*, *Atractylodis Macrocephalae Rhizoma* and *Saposhnikovia Radix*, regulates the release of cytokines in murine macrophages.** *PLoS One* 2013; 8(11):e78622.

49. Du CY, Choi RC, Dong TT, Lau DT, Tsim KW. **Yu Ping Feng San, an ancient Chinese herbal decoction, regulates the expression of inducible nitric oxide synthase and cyclooxygenase-2 and the activity of intestinal alkaline phosphatase in cultures.** *PLoS One* 2014; 9(6):e100382.

50. Lu J, Yu Y, Wang XJ, Chai RP, Lyu XK, Deng MH, et al. **Mechanism of Shengmai Injection () on Anti-Sepsis and Protective Activities of Intestinal Mucosal Barrier in Mice.** *Chin J Integr Med* 2021.

51. Zhou E, Li Y, Wei Z, Fu Y, Lei H, Zhang N, et al. **Schisantherin A protects lipopolysaccharide-induced acute respiratory distress syndrome in mice through inhibiting NF-kappaB and MAPKs signaling pathways.** *Int Immunopharmacol* 2014; 22(1):133-140.

52. Minami M, Konishi T, Makino T. **Effect of Hochuekkito (Buzhongyiqitang) on Nasal Cavity Colonization of Methicillin-Resistant Staphylococcus aureus in Murine Model.** *Medicines (Basel)* 2018; 5(3).
53. Kitahara M, Takayama S, Akaishi T, Kikuchi A, Ishii T. **Hochuekkito can Prevent the Colonization of Methicillin-Resistant Staphylococcus aureus in Upper Respiratory Tract of Acute Stroke Patients.** *Front Pharmacol* 2021; 12:683171.
54. Matsumoto T, Noguchi M, Hayashi O, Makino K, Yamada H. **Hochuekkito, a Kampo (traditional Japanese herbal) Medicine, Enhances Mucosal IgA Antibody Response in Mice Immunized with Antigen-entrapped Biodegradable Microparticles.** *Evid Based Complement Alternat Med* 2010; 7(1):69-77.