

TITLE PAGE:

TITLE: Secondary Dengue Infection Elicits Earlier Elevations in IL-6 and IL-10 Levels.

Running title: High IL-6 and IL-10 in dengue infection

Authors:

Espindola Sonia L ^{1, 5}, Fay Jessica ^{1, 5}, Carballo Graciela M ², Pereson Matías J ^{3, 5}, Aloisi Natalia ⁶,
Badano María Noel ^{4, 6}, Ferreras Julián ^{1, 5}, Argüelles Carina ¹, Pezzarini Simón ¹, Chuit Roberto ⁷,
Miretti Marcos ^{1, 5}, Di Lello Federico A ^{3, 5}, and Baré Patricia ^{4, 6}.

Affiliations:

¹Laboratorio GIGA, Instituto de Biología Subtropical (IBS), Facultad de Ciencias Exactas
Químicas y Naturales, Universidad Nacional de Misiones (UNaM), Consejo Nacional de
Investigaciones Científicas y Técnicas (CONICET), Misiones, Argentina

²Laboratorios CEBAC SRL, Posadas, Misiones, Argentina

³Universidad de Buenos Aires. Facultad de Farmacia y Bioquímica. Instituto de Investigaciones
en Bacteriología y Virología Molecular (IBaViM). Buenos Aires, Argentina.

⁴Instituto de Medicina Experimental (IMEX), Consejo Nacional de Investigaciones Científicas y
Técnicas (CONICET), Academia Nacional de Medicina, Buenos Aires, Argentina

⁵Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET), Buenos Aires, Argentina

⁶Instituto de Investigaciones Hematológicas (IIHEMA), Academia Nacional de Medicina.

⁷Instituto de Investigaciones Epidemiológicas (IIE), Academia Nacional de Medicina.

Corresponding author: Patricia Baré **E-mail:** patobare@gmail.com;

pbare@hematologia.anm.edu.ar

Keywords: Dengue virus, Cytokines, infection phases, secondary infection, febrile stage

Address: Academia Nacional de Medicina, Pacheco de Melo 3081, 1425, CABA, Buenos Aires;
and, Facultad de Ciencias Exactas Químicas y Naturales, Universidad Nacional de Misiones
(UNaM), Félix de Azara 1552, Posadas, Misiones, Argentina.

Phone: +54 48091000 / +5403764435099

Fax: +5403764425414

ABSTRACT:

Introduction: Dengue virus (DENV) represents a global health concern. Symptomatic infection causes a wide range of clinical manifestations, from mild dengue fever to severe disease, characterized by vascular permeability and bleeding. Previous reports indicated that the exacerbated expression of some cytokines is implicated in the progression of the disease. However, their time of expression within the infectious period remain uncertain. The aim of this study was to assess IL-6 and IL-10 level kinetics distinguishing two phases within the febrile stage in DENV infected patients.

Methods: We conducted a retrospective study on samples from 2016 and 2020 DENV outbreaks in Argentina. Viremic patients were categorized in Phase I and II, based on anti-DENV IgM presence. Cytokine levels, clinical parameters, and type of infection were analyzed.

Results: Our analysis included samples from 259 patients in the febrile stage. Of these, 184 patients (71%) were classified into Phase I, while 75 patients (29%) were in Phase II. Ninety-nine patients showed secondary infection (38.2%). Notably, secondary infections exhibited earlier IL-6 and IL-10 elevation than primary infections, suggesting pre-existing immune memory priming the immune response. Thrombocytopenia and elevated AST were associated with Phase II, secondary infection, and hospitalization. Elevation of IL-6 and IL-10 correlated with low platelet counts, suggesting their association with clinical manifestations.

Conclusion: In cases of secondary dengue infections, cytokine levels increase at an earlier stage of the disease compared to primary infections, where elevated cytokine levels are typically observed during the later febrile phase. Further research with broader cytokine panels is warranted to validate these findings.

INTRODUCTION

Dengue is a human disease caused by the infection with any one of four genetically related dengue virus serotypes (DENV-1, -2, -3 and -4), which are transmitted to individuals through the bite of infected mosquitoes belonging to the *Aedes* species. Multiple factors have contributed to the reemergence of DENV infections as a significant global public health concern during the past 2 decades¹.

DENV infection can vary in presentation, ranging from asymptomatic cases or mild illness to severe disease. Severe dengue is characterized by vascular permeability, plasma leakage, massive bleeding, and, in some cases, liver compromise, organ impairment, and even death ¹².

In patients who experience a sudden deterioration of symptoms during the critical phase, which typically occurs around 3-7 days after the onset of illness, close monitoring is essential to prevent complications and reduce the risk of mortality ².

The known contributors behind severe and fatal cases of dengue are viral serotype ³, a second heterotypic infection which could cause an antibody-dependent enhancement (ADE) ^{4,5} and different aspects of the host innate and adaptive immune response ⁶. Cytokines, mediators released because of complex interactions between DENV and host immune responses, have been implicated to play a role in the progression of severe dengue disease ^{7,8}. Excessive generation of pro-inflammatory cytokines, such as and IL-6 proved to contribute to the production of antiplatelet or anti endothelial cell antibodies, which results in a deficiency in coagulation, leading bleeding with dengue infection ⁹⁻¹¹. Conversely, the presence of anti-inflammatory cytokines such as IL-10 led to compromised immune clearance and persistent infectious effects during acute viral infection ¹²⁻¹⁴.

The current application of these potential markers remains uncertain by the limited understanding of how their expression evolves during the initial phases of the infection and whether their increase is protective or detrimental to the disease outcome.

Because previous reports indicated that the events triggering the release of inflammatory mediators take place very early in illness (specifically, initial period of the febrile phase)¹⁵ and in line with the CDC's description of the febrile phase as having a biphasic nature, our study introduces a classification of two sub-phases within the febrile phase. In this context, we examined the levels of IL-6 and IL-10 to analyze their kinetics in patients with DENV across these two distinct febrile phases.

MATERIALS AND METHODS

Patients and study design

This is a retrospective study carried out on archived plasma/serum samples collected from patients in the city of Posadas, Misiones, Argentina, referred for diagnosis of dengue infection, during 2 consecutive dengue epidemic periods, 2016 and 2020. A group of 259 samples within the febrile stage, showing positive viremia (PCR-RNA+) or DENV NS1 antigen (+) were randomly selected and included in our analyses. Samples were stored at -80°C until used for cytokine and serotype assays.

Dengue diagnosis and laboratory studies

DENV diagnostic was carried out by immunochromatographic test, RDT, (SD BIOLINE Dengue DUO kit, Abbott, USA) for DENV NS1 antigen detection or the RealStar® Dengue RT-PCR Kit 3.0 (Altona Diagnostics, Hamburg, Germany) for virus genome detection. Specific antibodies to DENV, IgM and IgG, were determined by ELISA (Dia.Pro Diagnostic Bioprobes s.r.l., Milan, Italy). DENV serotypes were determined using the CDC DENV 1–4 real time PCR assay modified by Santiago et al.¹⁶.

Routine lab determinations were performed, and parameters associated with bleeding were assessed in our cohort. Furthermore, we analyzed the concentration of hepatic enzymes, ALT (alanine transaminase) and AST (aspartate aminotransferase), as well as total protein, albumin,

and bilirubin. Certain parameters were categorized based on specific thresholds that were previously associated with dengue severity^{15,17}. For instance, white blood cell counts were considered indicative of severity when they fell below 5,000/ μ L; similarly, platelet counts below 100,000/ μ L. Liver enzyme concentration exceeding the upper limit of the normal reference range (40 UI/mL) were considered elevated.

Classification of phases and type of infection

Patients within the febrile stage were categorized into two phases: the early phase (Ph I), characterized by the presence of DENV-RNA (+) or NS1-Ag (+), and the late phase (Ph II), characterized by the presence of viremia along with anti-DENV IgM. The type of infection was defined based on the presence or absence of anti-DENV IgG antibodies. Secondary infections were determined when NS1 antigen or viral genome were present concurrently with anti-DENV IgG antibodies, and patients with viremia but without anti-DENV IgG antibodies were categorized as primary infections, irrespective of the presence of anti-DENV IgM.

IL-6 and IL-10 determination

The concentrations of IL-6 (standard curve range: 0 - 300 pg/mL) and IL-10 (standard curve range: 0 - 500 pg/mL) were determined using commercial reagents based on enzyme linked immunosorbent assay (ELISA) (BD-Biosciences, San Diego, California, United States). Procedures were carried out following the supplier's recommendations. Duplicates were performed in selected samples to verify the accuracy of the results. A control group comprising 50 healthy blood donors, testing negative for dengue antibodies was included to establish cytokine' reference ranges.

Statistical Analysis

Continuous variables were compared using either the student's t-test or the Mann-Whitney U test. Categorical variables were assessed using the Chi-square test or Fisher's exact test. Confidence intervals were set at 95% (CI95) and a p value <0.05 was considered statistically significant. Statistical analysis was conducted using the SPSS statistical software package release 23.0 (IBM SPSS Inc., Chicago, IL, USA) and graphical representations were generated using GraphPad Prism 10.0.0 software.

Ethical Aspects

Patients included in the study were given and signed an informed consent. The experimental protocols and procedures carried out in this work were approved by the Biosafety Review Board, Ethical Committee of the Academia Nacional de Medicina, Buenos Aires (CEIANM) and the Ethical Committee of the Investigación Provincial, Misiones (CEIP).

RESULTS

Study Population

Sera or plasma samples from 259 patients, referred for dengue infection diagnosis during two dengue epidemic periods, were included in this study. Of these 128 (49.4%) samples were from the 2016 outbreak, and 131 (50.6%) were from the 2020 outbreak. Among the patients, 137 (52.9%) were female with a median age (Q1-Q3) of 44 years (29-63). Our analysis of 205 serotypes revealed a prevalence of 86.3% (n=177) for DENV-1 and 13.7% (n=28) for DENV-4. Additionally, 160 (61.8%) had primary infection, while the remaining 99 patients (38.2%) exhibited evidence of secondary infection. Notably, our study revealed a significant association between secondary infections and a higher incidence of individuals with platelet counts below 100,000/ μ l ($p=0.024$) compared to primary infections. Specifically, 12 out of 160 (7.7%) patients with primary infections and 17 out of 99 (17.7%) with secondary infection presented platelet counts lower than 100,000/ μ L.

Characteristics of Clinical and Immunological Parameters within the febrile stage

Within the febrile stage, we found 184 patients (71.0%) in Ph I, and 75 patients (29%) in Ph II.

Gender differences were not observed ($p=0.891$); however, patients in Ph I were, on average, 10 years younger than those in Ph II [41 years (29-60) vs. 51 (29-72), respectively, $p=0.031$].

Furthermore, as shown in Table 1, significant differences were also noted in DENV serotype

($p=0.007$) and the type of infection ($p<0.001$). Analyzing clinical parameters and cytokines

between the early and late phases, a significant increase in the number of individuals with

platelet counts below 100,000/ μL ($p=0.007$) in Ph II compared to Ph I was observed.

Additionally, a higher proportion of patients in Ph II had elevated AST values compared to

those in Ph I. Concerning the level of inflammatory and anti-inflammatory cytokines, only IL-6

exhibited higher concentrations during Ph I [7.67pg/mL for Ph I vs. 4.78pg/mL for Ph II,

$p=0.045$] while no significant differences were observed for IL-10 ($p=0.611$). Table 1 presents

all the epidemiological and clinical characteristics analyzed according to the phase within the

febrile stage.

The analysis of cytokine levels in relation to the febrile stage and the type of infection revealed

that IL-6 and IL-10 concentrations ($p=0.005$ and $p<0.001$, respectively) were elevated in Ph I

from secondary infections, compared to Ph I in primary infections. Conversely, primary

infections showed higher values in Ph II than that of secondary infections, for both cytokines

(IL-6, $p=0.009$; IL-10, $p=0.003$). Table 2 provides details on cytokine levels at different febrile

phases and types of infection. To establish a reference, levels of IL-6 and IL-10 were also

determined in healthy individuals ($n=50$) demonstrating higher levels in DENV-infected

individuals ($n=259$). Specifically, IL-6 concentration was 3.48 pg/mL in healthy controls and

6.71 pg/mL in DENV-infected patients ($p<0.001$) while for IL-10, levels were 5.75 pg/mL in

healthy individuals and 16.73 pg/mL for those infected with DENV ($p<0.001$) (Figure 1).

As a final assessment, an analysis to evaluate the relationship between clinical parameters and cytokine concentration during Ph II of secondary infections was performed. In summary, the analysis revealed that elevated IL-6 levels were associated with low platelet counts and tended to be associated with high ALT. On the other hand, elevated IL-10 levels were associated with WBC >5000, low platelets, and increased AST (Table 3).

Characteristics of hospitalized patients

Hospitalization often indicates greater disease severity in dengue infection. Since 26 patients (10%) required hospitalization due to DENV infection complications, we analyzed their clinical and immunological parameters. Our study revealed that hospitalized patients were, on average, 15 years older than outpatients ($p=0.007$). Among them, a higher proportion exhibited platelet values below 100,000/ μ L compared to outpatients ($p<0.001$). Additionally, hospitalized patients displayed elevated IL-6 and AST concentrations, highlighting the severity of the condition ($p<0.001$ and $p=0.047$, respectively). Table 4 summarizes the epidemiological and clinical characteristics of DENV infected patients based on hospitalization.

DISCUSSION

IL-6 and IL-10 are two cytokines known to play critical roles in the pathogenesis of DENV infection^{18,19}. Their levels can vary depending on the type of infection and the severity of the disease, warranting further research to better understand their significance in disease progression since a persistent controversy has been observed^{14,20}. Recent studies have suggested that the increase in cytokine levels occurs in very early stages, triggered by the innate immune response following virus entry. To investigate this, we divided the febrile viremic stage into two phases and examined the kinetic of IL-6 and IL-10 levels. One of our most notable findings is that in secondary dengue infections, cytokine levels rise earlier in the disease course compared to primary infections, in which elevated cytokine values

are detected during the later febrile phase. This suggests that in secondary infections, the immune system, guided by pre-existing immune memory, is primed for a more rapid response to the virus, leading to an anticipated immune response. However, the mechanisms governing differential clinical outcomes remain poorly defined.

Thrombocytopenia (platelet count <100,000/ μ L) is a hallmark of dengue hemorrhagic fever, and it remains one of the current criteria for diagnosing the condition ²¹. We observed a significant increase in the individuals with low platelet counts, and elevated AST values in Ph II. Focusing on the very early stages of the disease, when laboratory parameters are still within normal ranges, may have limited our ability to observe further significant clinical alterations. Additionally, patients in Ph II were older than those in Ph I, suggesting age may impact disease outcomes, although this factor cannot be conclusively rule out.

Our study also identified a higher incidence of individuals with platelet count below 100,000/ μ L in cases of secondary infections, consistent with previous reports in which the risk of severe dengue increases in patients with heterotypic secondary DENV infection ^{22,23}. However, it is essential to note that when sequential DENV infections are closely spaced, significant cross-protection may occur, potentially altering the association between type of infection and severity ²⁴. Remarkably, it is important to mention that DENV-1 was the dominant serotype in both consecutive outbreaks analyzed here, with the occurrence of DENV-4 in a lower percentage (30%) of the cases of 2020, a pattern consistent with previous publications ^{25,26}. However, a limitation of our study is the lack of comprehensive information on all serotypes, and the inability to determine whether secondary infections within our population were heterotypic or homotypic.

It is worth emphasizing that cytokine values for healthy controls were similar to that observed for healthy populations in previous reports ^{27,28} but lower than the levels exhibited in the group of infected patients, as shown in Figure 1.

Although reports of elevated IL-6 levels in early acute phase in bleeding patients were published ¹¹, others observed significantly higher IL-6 levels associated with hemorrhagic fever only in patients infected with DENV-2 but not with DENV-1 ²⁹. On the other hand, high levels of IL-10 in dengue during early illness were indicator of an altered antiviral response and this association was particularly observed in patients who progressed to dengue hemorrhagic fever ^{30,31}.

While direct correlation between cytokine levels in the acute phase and the severity of the disease was not individually evaluated in this study, the analysis of hospitalized patients (Table 4) revealed an alteration in AST concentrations and platelets, combined with elevated levels of IL-6, highlighting the role of these three parameters as warning signs in hospitalized individuals. Additionally, the older median age in the hospitalized group compared to outpatients may have contributed to the increased risk in these patients. It is essential to note that cytokine elevation does not necessarily correlate with a worsened prognosis since cytokines play a vital role in pathogen eradication ³². However, in conjunction with other risk factors, an exacerbated cytokine response may contribute to disease progression ³³.

In Ph II of secondary infections, our analysis revealed the association between clinical parameters related to dengue severity and the elevation of IL6 and IL10. Based on these findings, it seems plausible that elevated IL6 and IL10 levels in Ph I might be indicative of deteriorating conditions in Ph II.

Study Limitations

While the study boasts several strengths, such as its large sample size, inclusion of patients from different outbreaks, and the separation of two febrile phases, it is essential to acknowledge its limitations. This is not a sequential longitudinal study; rather samples from different phases correspond to distinct individuals, each at a specific point in their infection. Nevertheless, they all belong to the same population in Posadas, Misiones, Argentina.

Moreover, the study focused on a limited set of cytokines, and further studies should explore the roles of other immune factors in dengue infection. Lastly, the small number of hospitalized patients in our study precluded the possibility of conducting a logistic regression analysis.

Conclusions

In summary, our study provides valuable insights into the kinetics of IL-6 and IL-10 in different febrile phases and types of infections. The association between particular cytokines and specific disease phases highlights the importance of categorizing patients upon DENV diagnosis based on the type of infection they have. Moreover, considering similar days since symptoms onset will facilitate more precise comparisons. The measurement of cytokine levels that might act as indicators for disease progression at this moment, could be an invaluable tool in making earlier decisions regarding patient interventions, especially in the context of secondary infections. Future studies should be conducted to validate our conclusions and explore the relationship between a broader panel of cytokine and dengue pathogenesis.

Data availability statement: The data that support the findings of this study are available from the corresponding author upon reasonable request.

Conflict of interest statement: On behalf of all authors, the corresponding author states that there is no conflict of interest.

Author contributions:

SLE, FAD and PB: Conceptualized and designed the study, collected, and validated data, performed statistical analysis and data interpretation, organized, and curated the dataset, wrote the initial draft of the manuscript and its revision, created and approved the final version to be submitted. MJP, AN, PS: conducted dengue diagnostic assays and cytokine ELISA assays, collected and validated data, reviewed and approved the final version of the manuscript to be submitted. CGM, AC, BMN, FJ, MM and CR: performed data analysis, organized, and curated the dataset, reviewed the article critically for important intellectual content, approved the final version of the manuscript to be submitted.

Funding: This work was supported by grant PIP N°1122017010 0781 CO from the National Council for Research and Technology (CONICET), Buenos Aires, Argentina, and funds provided by CEBAC Laboratories SRL, Posadas, Misiones, Argentina.

Acknowledgements

MJP, FJ, MM, SLE, FAD and PB are members of the National Research Council (CONICET). Some aspects of this investigation could not have been fulfilled without the generous financial support of the Fundación René Baron.

Ethics statement: The study was conducted in accordance with the Declaration of Helsinki and approved by the Institutional Ethics Committee of the Academia Nacional de Medicina, Buenos Aires (TI N°13157/19/X) and the Ethical Committee of the Investigación Provincial, Misiones (CEIP). Informed consent was obtained from all the individuals.

REFERENCES

1. WHO. Dengue and Severe Dengue, Fact Sheets, World Health Organization (2020). Available at: <https://www.who.int/news-room/fact-sheets/detail/Dengue-and-severe-Dengue>.
2. Centers for Disease Control and Prevention, National Center for Emerging and Zoonotic Infectious Diseases (NCEZID), Division of Vector-Borne Diseases (DVBD) Last Reviewed: April 13, 2023.
3. Hubert B. and Halstead S.B. Dengue 1 virus and dengue hemorrhagic fever, French Polynesia, 2001. *Emerg Infect Dis* 15, 1265–1270 (2009).
4. Guzman, M. G., Alvarez M. and Halstead S.B. Secondary infection as a risk factor for dengue hemorrhagic fever/dengue shock syndrome: an historical perspective and role of antibody-dependent enhancement of infection. *Arch Virol* 158, 1445–1459 (2013).
5. Katzelnick L. C., Lionel G., Elizabeth, H. M., Carlos M. J. and Kuan G. Antibody-dependent enhancement of severe dengue disease in humans. 6836, 1–12 (2017).
6. Wijeratne D., Samitha F., Laksiri G., Chandima J., Anushka G., Samarasekara S., Wijewickrama A., Hardman C., Graham O., Malavige G.N. Quantification of dengue virus specific T cell responses and correlation with viral load and clinical disease severity in acute dengue infection. *PLoS Negl Trop Dis* 12, 1–16 (2018).
7. Srikiatkachorn A., Mathew A., and Rothman A. Immune Mediated Cytokine Storm and Its Role in Severe Dengue. *Semin Immunopathol.* 176, 139–148 (2017).
8. Patro A.R.K, Mohanty S., Prusty B.K., Singh D.K., Gaikwad S., Saswat T., Chattopadhyay S., Das B.K., Tripathy R., and Ravindran B. Cytokine signature associated with disease severity in dengue. *Viruses* 11, 1–12 (2019).
9. Kamaladasa A., Gomes L., Wijesinghe A., Jeewandara C., Ying Xiu Toh, Jayathilaka D., Graham S.O., Katja F., and Malavige G.N. Altered monocyte response to the dengue virus in those with varying severity of past dengue infection. *Antiviral Res* 169, 104554 (2019).
10. Rachman, A. & Rinaldi, I. Coagulopathy in dengue infection and the role of interleukin-6. *Acta Med Indones* 38, 105–108 (2006).
11. Imad, H. A., Phumratanaprapin W., Phonrat B., Chotivanich K., Charunwatthana P., Muangnoicharoen S., Khusmith S., Tantawichien T., Phadungsombat J., Nakayama E., Konishi E., and Shioda T. Cytokine Expression in Dengue Fever and Dengue Hemorrhagic Fever Patients with Bleeding and Severe Hepatitis. *Am J Trop Med Hyg* 102, 943–950 (2020).
12. Puc I., Ho T.C., Yen K.L., Vats A., Tsai J.J., Chen P.L., Chien Y.W., Lo Y.C., and Perng G.C. Cytokine Signature of Dengue Patients at Different Severity of the Disease. *Int J Mol Sci* 22, (2021).
13. Nwe M. K., Ngwe Tun M., Win Myat T., Sheng Ng C., Htun M., Htin Lin, Hom N.S., Min Soe A., Ngono A. E., Hamano S., Morita K., Zin Thant K., Shresta S., Myat Thu H., and Moi M.L. Acute-phase Serum Cytokine Levels and Correlation with Clinical Outcomes in

334 Children and Adults with Primary and Secondary Dengue Virus Infection in Myanmar
335 between 2017 and 2019. *Pathogens* 11, (2022).

336 14. Friberg H., Beaumier C., Park S., Pazoles P., Endy T.P., Mathew A., Currier J., Jarman R.,
337 Anderson K. B., Hatch S., Thomas J.S., and Rothman L.A. Protective versus pathologic
338 pre-exposure cytokine profiles in dengue virus infection. *PLoS Negl Trop Dis* 12, 1–15
339 (2018).

340 15. Pongpan, S., Patumanond, J., Wisitwong, A., Tawichasri, C. & Namwongprom, S.
341 Validation of dengue infection severity score. *Risk Manag Healthc Policy* 7, 45–49
342 (2014).

343 16. Santiago G., Vergne E., Quiles Y., Cosme J., Juan Medina J.V., Medina F., Colón C.,
344 Margolis H., and Jorge L Muñoz-Jordán. Analytical and Clinical Performance of the CDC
345 Real Time RT-PCR Assay for Detection and Typing of Dengue Virus. *PLoS Neglected*
346 *Tropical Diseases* vol. 7 Preprint at <https://doi.org/10.1371/journal.pntd.0002311>
347 (2013).

348 17. Sangkaew S., Ming D., Boonyasiri A., Honeyford K., Kalayanarooj S., Yacoub S., Dorigatti
349 I., and Holmes A. Risk predictors of progression to severe disease during the febrile
350 phase of dengue: a systematic review and meta-analysis. *Lancet Infect Dis* 21, 1014–
351 1026 (2021).

352 18. Tran L., Radwan I., Nhat Minh L., Khai Low S., Hashan M.R., Gomaa M.D., Abdelmongy
353 M., Abdelaziz A., Mohamed A., Tawfik G., Mizukami S., Hirayama K., and Huy N.T. Role
354 of cytokines produced by T helper immune-modulators in dengue pathogenesis: A
355 systematic review and meta-analysis. *Acta Trop* 216, 105823 (2021).

356 19. Tsung-Ting Tsai, Yi-Jui Chuang, Yee-Shin Lin, Shu-Wen Wan, Chia-Ling Chen, Chiou-Feng
357 Lin. An emerging role for the anti-inflammatory cytokine interleukin-10 in dengue virus
358 infection. *J Biomed Sci* 20, 40 (2013).

359 20. Rathakrishnan A., Wang S.M., Hu Y., Khan A.M., Ponnampalavanar S., Lum L.C.,
360 Manikam R., and Sekaran S.D. Cytokine Expression Profile of Dengue Patients at
361 Different Phases of Illness. *PLoS One* 7, 1–10 (2012).

362 21. Zarco, R. M., Espiritu Campos, L. and Chan, V. Preliminary report on laboratory studies
363 on Philippine hemorrhagic fever. *J Philipp Med Assoc* 33, 676–683 (1957).

364 22. Sierra B., Perez A.B., Vogt K., Garcia G., Schmolke K., Aguirre E., Alvarez M., Kern F.,
365 Kourí G., Hans-Dieter Volk, and Maria G Guzman. Secondary heterologous dengue
366 infection risk: Disequilibrium between immune regulation and inflammation? *Cell*
367 *Immunol* 262, 134–140 (2010).

368 23. Tsheten T., Clements A.C.A., Gray D.J., Adhikary R.K., Furuya-Kanamori L., and Wangdi
369 K. Clinical predictors of severe dengue: a systematic review and meta-analysis. *Infect*
370 *Dis Poverty* 10, 123 (2021).

371 24. Anderson B.K., Gibbons V.R., Cummings D., Nisalak A., Green S., Libraty D. H., Jarman
372 R.G., Srikiatkachorn A., Mammen M.P., Darunee B., In-Kyu Yoon, and Endy T. A shorter
373 time interval between first and second dengue infections is associated with protection
374 from clinical illness in a school-based cohort in Thailand. *J Infect Dis* 209, 360–368
375 (2014).

25. Flichman M.D., Pereson M.J., Baré P., Espindola S.L., Carballo G.M., Albrecht A., Agote F., Alter A., Bartoli S., Blanco S., Blejer J., Borda M., Bouzon N., Carrizo H.J., Etcheverry L., Fernandez R., Figueroa Reyes M.I., Gallego S., Hahn R., Luna S.G., Marranzino G., Suarez Romanazzi J., Rossi A., Troffe A., Chang-Chi Lin, Martínez A.P., García G., and Di Lello F.A. Epidemiology of dengue in Argentina: antibodies seroprevalence in blood donors and circulating serotypes. *Journal of Clinical Virology* 147, 105078 (2022).
26. Epidemiol, S. Boletín Integrado de Vigilancia SE19/2020. 1–47 (2020).
27. Said E.A., Al-Reesi I., Al-Shizawi N., Jaju S., Al-Balushi M.S., Koh C.Y., Al-Jabri A.A., and Jeyaseelan L. Defining IL-6 levels in healthy individuals: A meta-analysis. *J Med Virol* 93, 3915–3924 (2021).
28. Sarris A.H., Kliche K.O., Pethambaram P., Preti A., Tucker S., C Jackow, Messina O., Pugh W., Hagemester F.B., McLaughlin P., Rodriguez M.A., Romaguera J., Fritsche H., Witzig T., Duvic M., Andreeff M., and Cabanillas F. Interleukin-10 levels are often elevated in serum of adults with Hodgkin’s disease and are associated with inferior failure-free survival. *Ann Oncol* 10, 433–440 (1999).
29. Cruz Hernández S.I., Puerta-Guardo H.N., Aguilar H.F., Mateos S.G., López Martinez I., Ortiz-Navarrete V., Ludert J.E., and Angel R.M. Primary dengue virus infections induce differential cytokine production in Mexican patients. *Mem Inst Oswaldo Cruz* 111, 161–167 (2016).
30. Dayarathna S., Jeewandara C., Gomes L., Somathilaka G., Jayathilaka D., Chandran V., Wijewickrama A., Narangoda E., Idampitiya D., Ogg S.G., and Malavige G.N. Similarities and differences between the ‘cytokine storms’ in acute dengue and COVID-19. *Sci Rep* 10, 19839 (2020).
31. Malavige G.N., Jeewandara C., Luckmaal Alles K.M., Salimi M., Gomes L., Kamaladasa A., Jayaratne S.D., and Ogg S.G. Suppression of virus specific immune responses by IL-10 in acute dengue infection. *PLoS Negl Trop Dis* 7, e2409 (2013).
32. Tuyen t.t., Thanh Viet n., Hang n.t., Giang n.t., Anh D.T, Hung H.V., Quyet D., Toan N.L., Cam T.D., and Van Tong H. Proinflammatory Cytokines Are Modulated in Vietnamese Patients with Dengue Fever. *Viral Immunol* 33, 514–520 (2020).
33. Wang W.H., Nayim A.U., Chang R.M., Assavalapsakul W., Po-Liang Lu, Chen Y.H., and Wang S.F. Dengue hemorrhagic fever – A systemic literature review of current perspectives on pathogenesis, prevention and control. *Journal of Microbiology, Immunology and Infection* 53, 963–978 (2020).

411 TABLES

412 Table 1. Epidemiological and Clinical characteristics by phase (n=259).

Characteristics	Total (n=259)	Phase 1 (n=184)	Phase 2 (n=75)	p
Age* (years)	44 (29-63)	41 (29-60)	51 (29-72)	0.031
Female gender (%)	137 (52.9)	98 (53.3)	39 (52)	0.891
Dengue serotype (%) [#]				
1	177 (86.3)	131 (82.9)	46 (97.9)	
4	28 (13.7)	27 (17.1)	1 (2.1)	0.007
Infection (%)				
Primary	160 (61.8)	133 (72.3)	27 (36.0)	
Secondary	99 (38.2)	51 (27.7)	48 (64.0)	<0.001
WBC <5000/ μ L (%)	170 (65.6)	119 (65.0)	51 (68.9)	0.663
Platelets <100,000/ μ L (%)	29 (11)	14 (7.8)	15 (20.8)	0.007
IL6 pg/mL	6.71 (3.4-15.2)	7.67 (3.5-15.8)	4.78 (3.2-10.7)	0.045
IL10 pg/mL	16.7 (9.8-41.1)	16.91 (10.1-40.2)	14.56 (8.4-63.2)	0.611
AST \geq ULN (U/L) [#] (%)	98 (37.8)	65 (41.7)	33 (61.1)	0.017
ALT \geq ULN (U/L) [#] (%)	63 (24.3)	45 (28.8)	18 (33.3)	0.606
Albumin g/dL [#]	4.1 (3.9-4.4)	4.21 (3.9-4.4)	4.04 (3.8-4.2)	0.005
Bilirubin mg/dL [#]	0.49 (0.34-0.73)	0.48 (0.34-0.73)	0.50 (0.34-0.74)	0.391
Total protein g/dL [#]	6.99 (6.7-7.3)	6.99 (6.7-7.4)	7.00 (6.7-7.2)	0.446

413 *Median (interquartile range), WBC: White blood cells, ULN: upper limit of normal (>40 U/L),

414 [#]Available in: Serotype 205 patients, ALT and AST 210 patients, Albumin 160 patients, Total
415 protein, and bilirubin 150 patients.

416

417

418 Table 2: Cytokine levels at different phases and type of infection (n=259).

Cytokines	Infection / Phase	Primary Infection	Secondary Infection	p by type
		(n=160)	(n=99)	of infection
		(n=133)	(n=51)	
	Phase I (n=184)	6.59 (3.3-14.5)	10.26 (5.5-26.8)	0.005
IL-6 pg/mL		(n=27)	(n=48)	
	Phase II (n=75)	7.7 (4.3-30.9)	4.28 (2.7-8.4)	0.009
p by phase		0.200	<0.001	
		(n=133)	(n=51)	
	Phase I (n=184)	14.30 (9.0-30.4)	36.46 (14.1-84.6)	<0.001
IL-10 pg/mL		(n=27)	(n=48)	
	Phase II (n=75)	30.34 (12.1-110.9)	11.25 (6.5-32.1)	0.003
p by phase		<0.001	<0.001	

419

420

421 **Table 3. Cytokine levels by clinical parameters during phase 2 of secondary infections (N=48)**

Characteristics	IL-6*	p	IL-10*	p
WBC 5000/ μ L				
<	4.3 (2.9-7.6)		14.0 (9.8-37.6)	
>	3.8 (2.3-10.7)	0.891	6.6 (5.7-11.2)	0.014
Platelets 100,000/ μ L				
<	10.7 (3.5-15.7)		42.3 (11.6-100.6)	
>	4.2 (2.6-6.2)	0.020	10.8 (6.2-18.0)	0.002
AST ULN (U/L) #				
<	3.3 (2.0-8.5)		7.6 (5.7-13.5)	
\geq	4.9 (3.5-10.7)	0.252	17.2 (11.3-42.3)	0.006
ALT ULN (U/L) #				
<	3.5 (2.8-6.3)		11.2 (6.3-20.5)	
\geq	6.2 (3.5-15.2)	0.082	17.2 (8.3-30.5)	0.330

422 *Median (interquartile range),
423 WBC: White blood cells, ULN: upper limit of normal (>40 U/L),
424 #Available in 34 patients.

425

426

427 **Table 4. Epidemiological and Clinical characteristics of DENV infected patients by**
428 **hospitalization (n=259).**

Characteristics	Outpatients (n=233)	Hospitalized (n=26)	p
Age* (years)	42 (28-62)	57 (40-74)	0.007
Female gender (%)	121 (51.9)	16 (61.5)	0.235
Infection			
Primary	145 (62.2)	15 (57.7)	0.401
Secondary	88 (37.8)	11 (42.3)	
WBC <5000/ μ L (%)	152 (65.8)	18 (69.2)	0.455
Platelets <100,000/ μ L	19 (8.4)	10 (40.0)	<0.001
IL6 pg/mL	6.21 (3.2-14.7)	11.07 (7.4-62.8)	<0.001
IL10 pg/mL	16.07 (9.4-40.8)	21.72 (11.5-62.6)	0.235
AST \geq ULN (U/L) #	83 (44.4)	15 (65.2)	0.047
ALT \geq ULN (U/L) #	56 (29.9)	7 (30.4)	0.566

429 *Median (interquartile range), WBC: White blood cells, ULN: upper limit of normal (>40 U/L),

430 #Available in: Serotype 205 patients, ALT and AST 210 patients.

431

432 **FIGURE LEGENDS:**

433 Figure 1: Cytokine values in DENV infected group (in black) and healthy population (in grey).

434 Median for IL-6, and IL-10 concentrations are expressed in log (pg/mL).

435